



WP5: Enzymatic refining of polysaccharides from Brown seaweed

Prof Gudmundur Hreggvidsson
Matis & University of Iceland

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MacroCascade Final Conference

WP5: Enzymatic refining for added-value products

Main bulk products from seaweeds are produced using **mechanical and/or physicochemical extraction/fractionation technologies.**

Seaweed meal

Hydrocolloids: alginate, carrageenan ..

Biocatalytic” tools”, enzymes and microbes, may offer new product possibilities and greater resource efficiency besides being environmental benign options to harsh physicochemical processes harmful to the environment.

Fermented seaweeds

Bioactive oligosaccharides,

Fermentable sugars,

Rare sugars,

Platform- and specialty chemicals,

Energy carriers (biofuels)

Possibilities are largely unexplored,



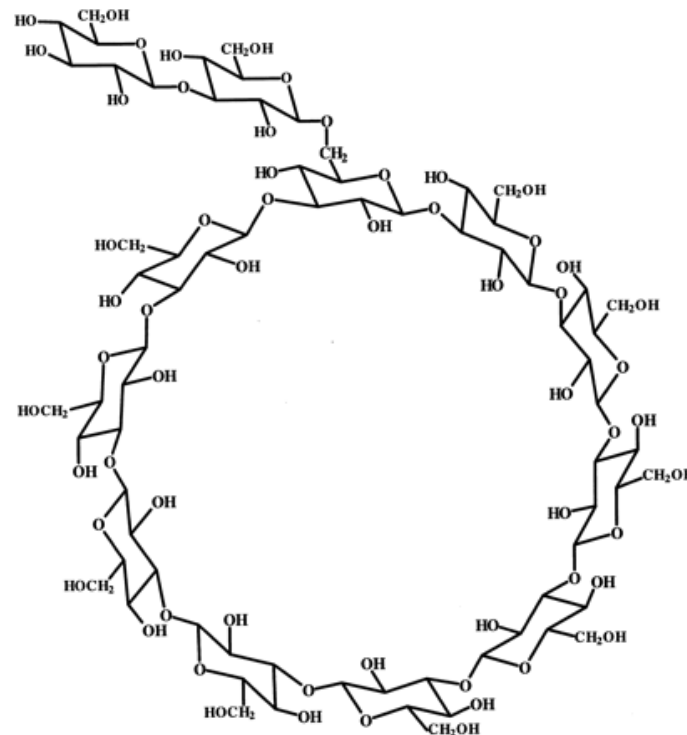
Enzymatic refining of polysaccharides from Brown macroalgae

Laminarin

A 1,3-(1,6)-beta-glucan

Aim: to use enzymes to make branched oligosaccharides enhanced in bioactivity

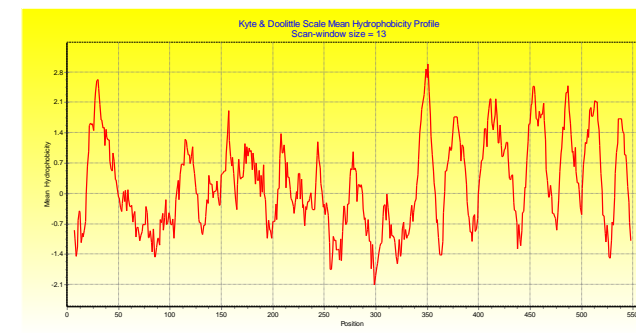
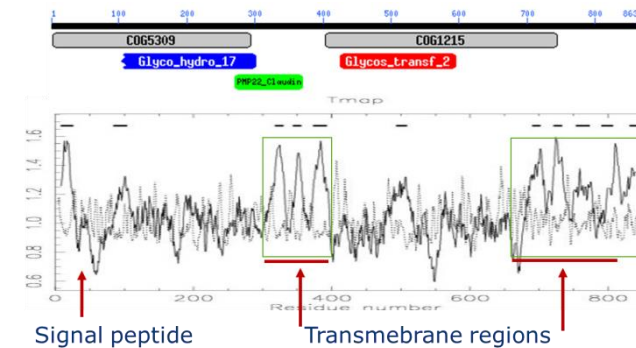
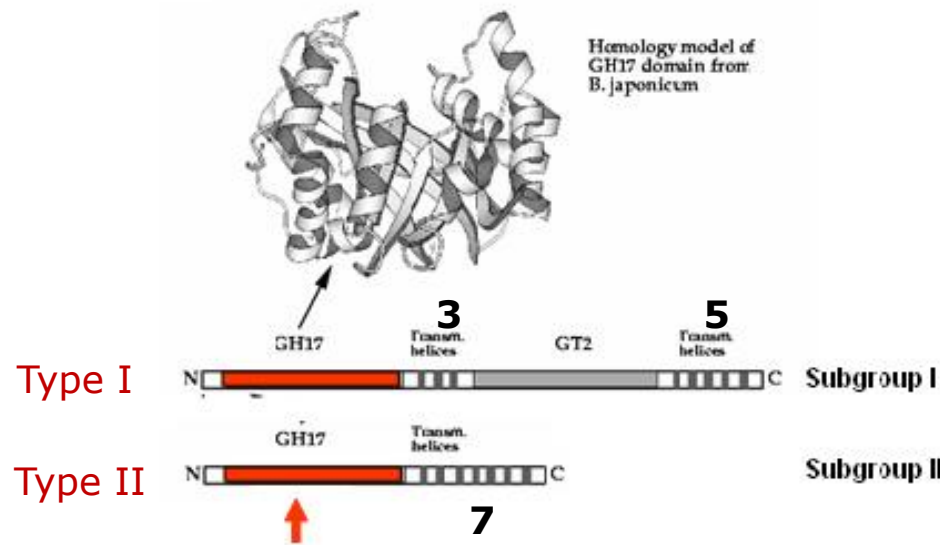
Investigated enzymes that could circularize beta-glucans in *Proeobacteria*



Membrane bound enzymes in Proteobacteria:

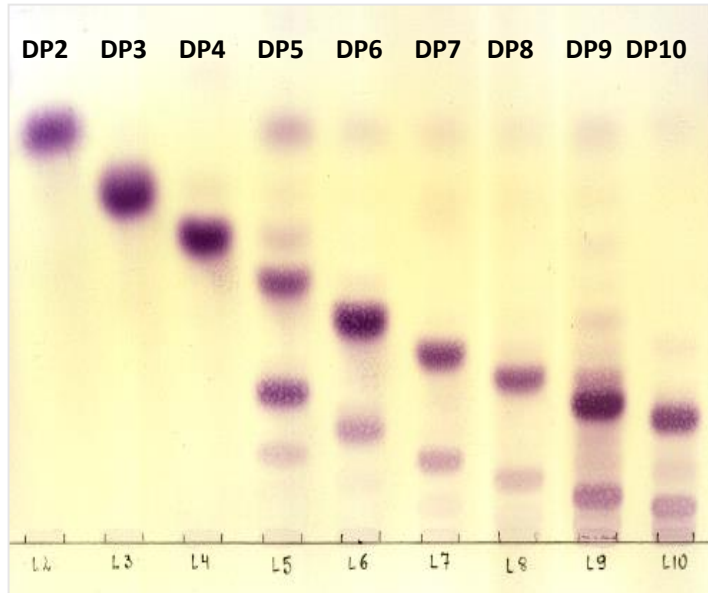
Type I: a two domain protein including a GH17 domain and a GT2 domain with two transmembrane regions.

Type II: a GH17 with one region: 7 transmembrane helices at the C terminal end.



Activity of the Type I GH17 catalytic domain

TLC analysis



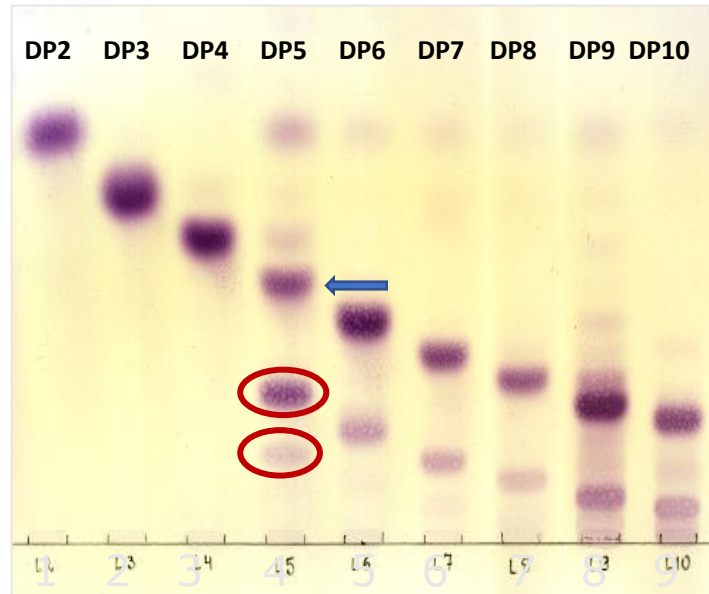
Laminari-oligosaccharides as substrates

- Lam-Glc₂
- Lam-Glc₃
- Lam-Glc₄
- Lam-Glc₅
- Lam-Glc₆
- Lam-Glc₇
- Lam-Glc₈
- Lam-Glc₉
- Lam-Glc₁₀



Activity of the Type I **GH17** catalytic domain

TLC analysis



Substrate

DP5

Products

DP8

(DP3 + DP5)

DP11

(DP3 + DP8)

Laminari-oligosaccharides
as substrates

Lam-Glc₂

Lam-Glc₃

Lam-Glc₄

Lam-Glc₅

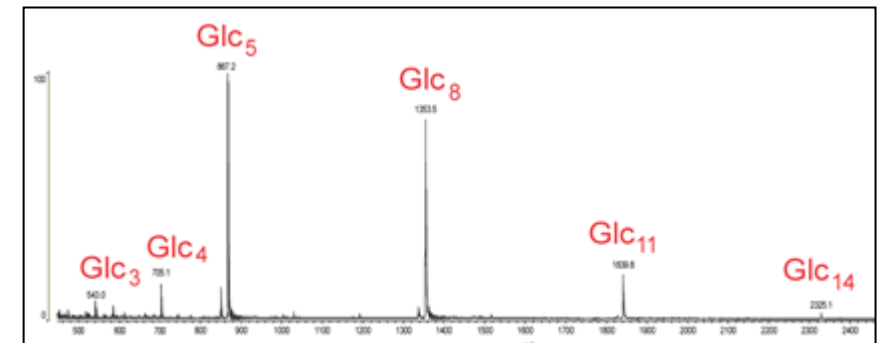
Lam-Glc₆

Lam-Glc₇

Lam-Glc₈

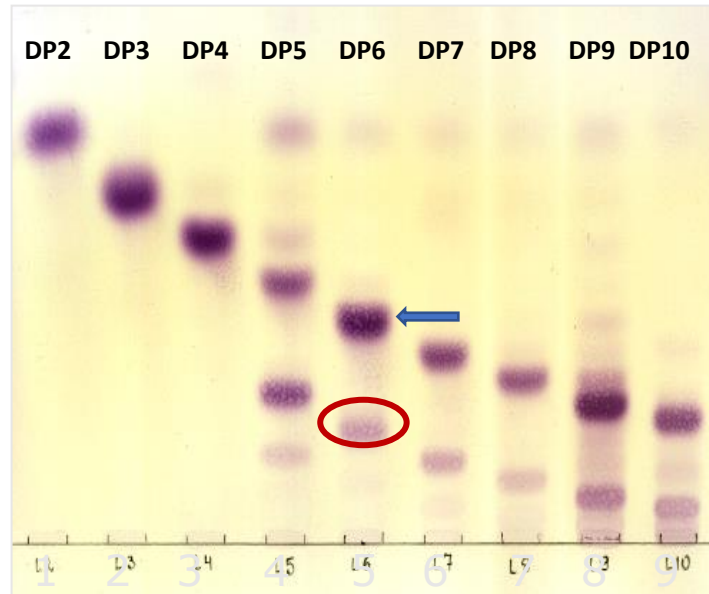
Lam-Glc₉

Lam-Glc₁₀



Activity of the Type I GH17 catalytic domain

TLC analysis



Laminari-oligosaccharides
as substrates

Lam-Glc2

Lam-Glc3

Lam-Glc4

Lam-Glc5

Lam-Glc6

Lam-Glc7

Lam-Glc8

Lam-Glc9

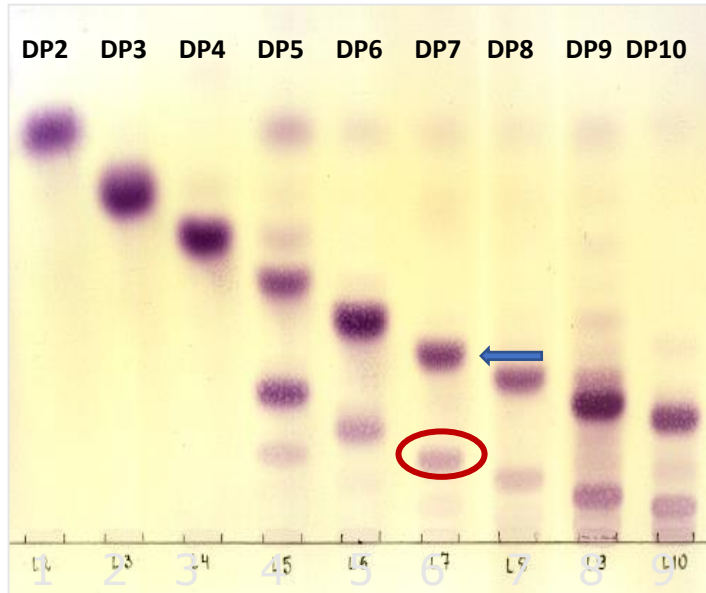
Lam-Glc10

Substrate	Products
DP6	DP10 (DP4 + DP6)
	DP14 (DP4 + DP10)



Activity of the Type I GH17 catalytic domain

TLC analysis



Laminari-oligosaccharides
as substrates

Lam-Glc2

Lam-Glc3

Lam-Glc4

Lam-Glc5

Lam-Glc6

Lam-Glc7

Lam-Glc8

Lam-Glc9

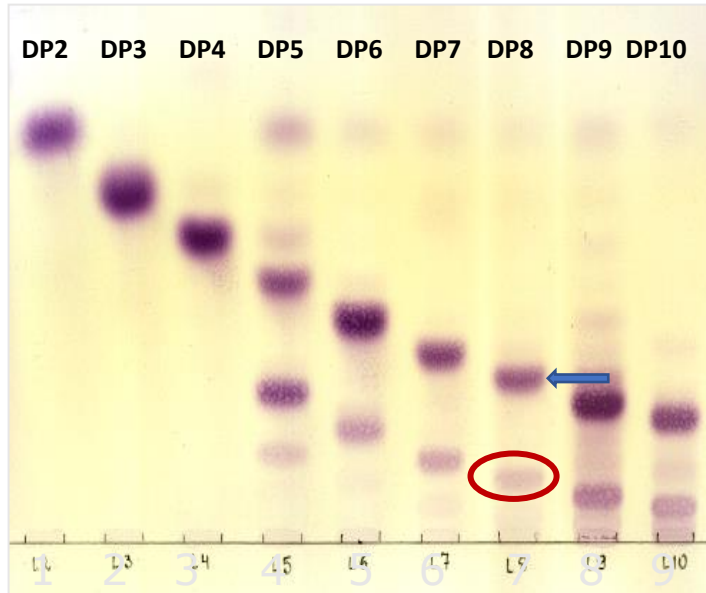
Lam-Glc10

Substrate	Products
DP7	DP12 (DP5 + DP7)
	DP17 (DP5 + DP12)



Activity of the Type I GH17 catalytic domain

TLC analysis



Laminari-oligosaccharides
as substrates

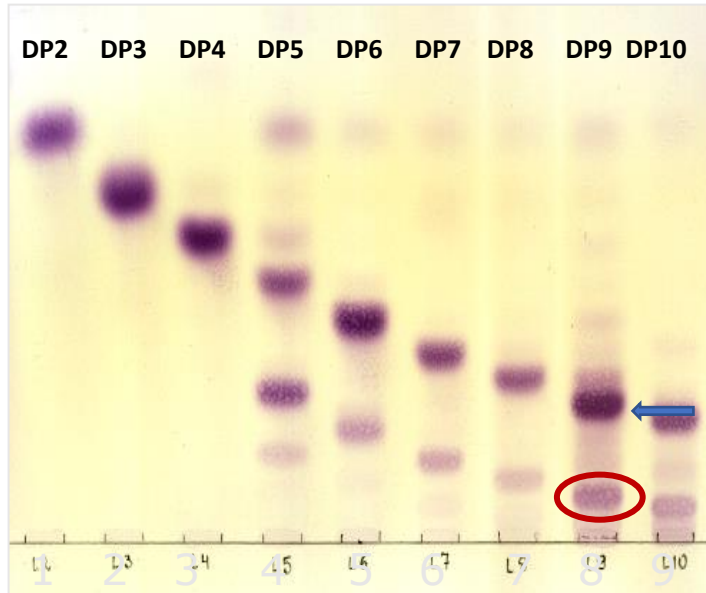
- Lam-Glc2
- Lam-Glc3
- Lam-Glc4
- Lam-Glc5
- Lam-Glc6
- Lam-Glc7
- Lam-Glc8**
- Lam-Glc9
- Lam-Glc10

Substrate	Products
DP8	DP14 (DP6 + DP8)
	DP20 (DP6 + DP14)



Activity of the Type I GH17 catalytic domain

TLC analysis



Laminari-oligosaccharides
as substrates

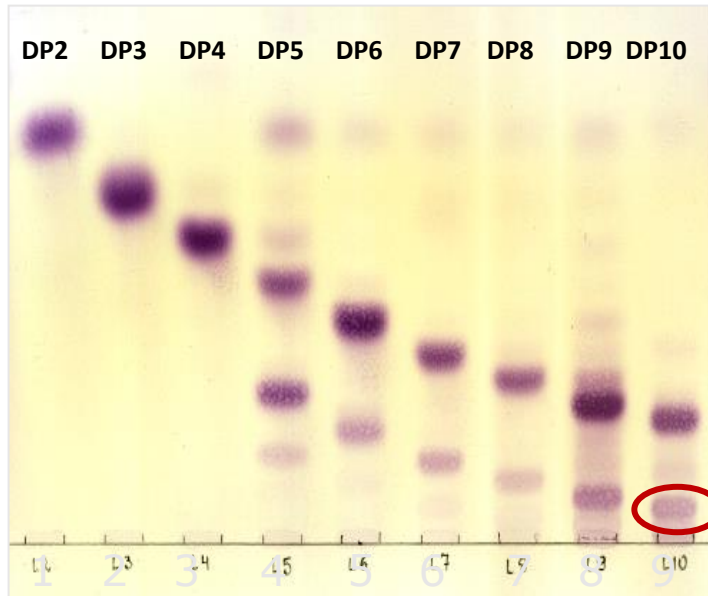
- Lam-Glc2
- Lam-Glc3
- Lam-Glc4
- Lam-Glc5
- Lam-Glc6
- Lam-Glc7
- Lam-Glc8
- Lam-Glc9**
- Lam-Glc10

Substrate	Product
DP9	DP16 (DP7 + DP9)
	DP26 (DP7 + DP16)



Activity of the Type I GH17 catalytic domain

TLC analysis



Laminari-oligosaccharides as substrates

- Lam-Glc2
- Lam-Glc3
- Lam-Glc4
- Lam-Glc5
- Lam-Glc6
- Lam-Glc7
- Lam-Glc8
- Lam-Glc9
- Lam-Glc10**

Substrate	Product
DP10	DP18 (DP8 + DP10)
	DP26 (DP8 + DP18)

Substrate	Product
DP5	DP8 (DP3 + DP5)
	DP11 (DP3 + DP8)

Substrate	Products
DP6	DP10 (DP4 + DP6)
	DP14 (DP4 + DP10)

Substrate	Product
DP7	DP12 (DP5 + DP7)
	DP17 (DP5 + DP12)

Substrate	Products
DP8	DP14 (DP6 + DP8)
	DP20 (DP6 + DP14)

Substrate	Product
DP9	DP16 (DP7 + DP9)
	DP26 (DP7 + DP16)

Substrate	Product
DP10	DP18 (DP8 + DP10)
	DP26 (DP8 + DP18)

Comparison of the activity GH17 domains from two different species

Glt20

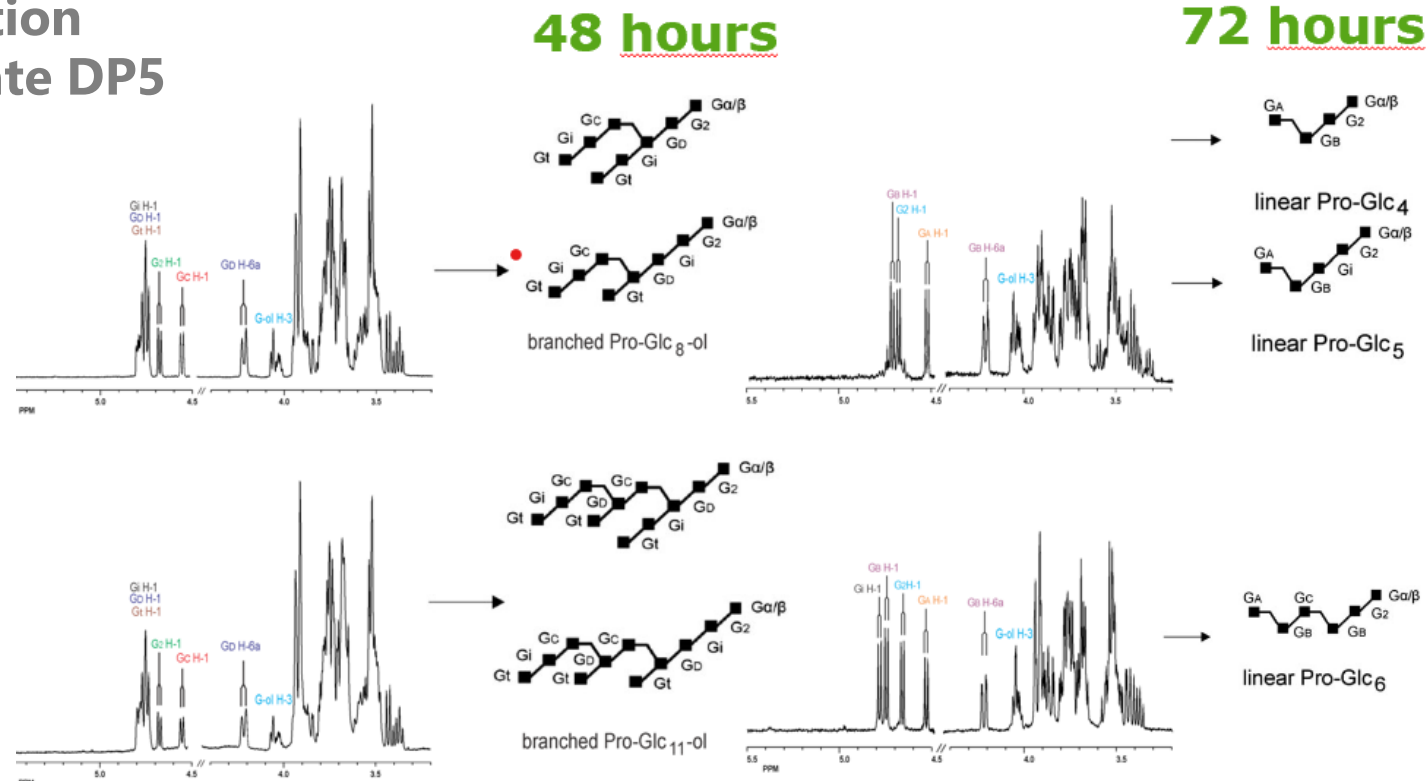
Substrate	Product
DP5	DP8 (DP3 + DP5) DP11 (DP3 + DP8)
Substrate DP6	Products DP10 (DP4 + DP6) DP14 (DP4 + DP10)
Substrate DP7	Product DP12 (DP5 + DP7) DP17 (DP5 + DP12)
Substrate DP8	Products DP14 (DP6 + DP8) DP20 (DP6 + DP14)
Substrate DP9	Product DP16 (DP7 + DP9) DP26 (DP7 + DP16)
Substrate DP10	Product DP18 (DP8 + DP10) DP26 (DP8 + DP18)

Glt7

Substrate	Product
<u>DP6</u>	DP9 (DP3 + DP6)
<u>DP7</u>	DP10 (DP3 + DP7)
Substrate <u>DP8</u>	Product DP11 (DP3 + DP8)
Substrate <u>DP9</u>	Product DP12 (DP3 + DP7)

What type of linkages are formed?

Incubation
Substrate DP5



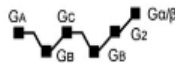
Substrate: Laminarioligosaccharide DP5 (both donor and acceptor)

When donor is depleted, hydrolysis occurs, and initially formed branched products are being degraded into linear kinked products

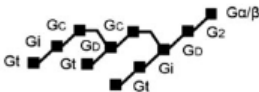
after prolonged reaction kinked linear products are formed **26% internal β-1,6-linkages** are observed

Enzyme toolbox for production of complex mixed linkage, kinked or highly branched, cyclic or linear β -glucans from laminarin polysaccharides, *in vitro*.

kinked

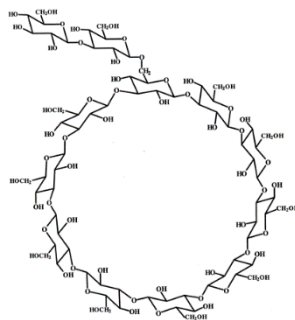


or branched



Matis has isolated and characterized a number of different transglucosidases active on beta-1,3 glucan polysaccharides such as laminarin.

or cyclic



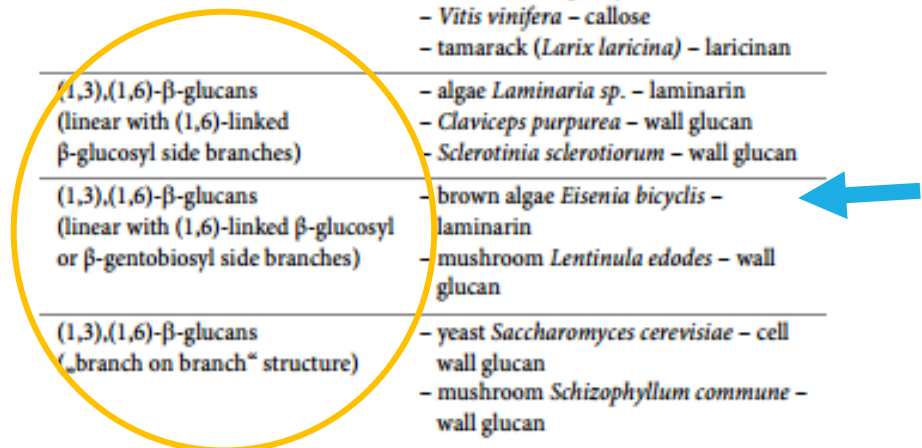
A patent has been acquired for the branching transglucosidase

Variety of beta- glucans having healthbenefits

Table 1. Examples of β -glucans with different structures, isolated from different natural sources (Stone and Clarke, 1992)

Type of β -glucan (structure description)	Natural source - trivial name of β -glucan
(1,3)- β -glucans (linear, homogeneous)	- bacterium <i>Alcaligenes faecalis</i> - curdlan - algae <i>Euglena gracilis</i> - paramylon - <i>Poria cocos</i> - pachyman - <i>Vitis vinifera</i> - callose - tamarack (<i>Larix laricina</i>) - laricinan
(1,3),(1,6)- β -glucans (linear with (1,6)-linked β -glucosyl side branches)	- algae <i>Laminaria sp.</i> - laminarin - <i>Claviceps purpurea</i> - wall glucan - <i>Sclerotinia sclerotiorum</i> - wall glucan
(1,3),(1,6)- β -glucans (linear with (1,6)-linked β -glucosyl or β -gentobiosyl side branches)	- brown algae <i>Eisenia bicyclis</i> - laminarin - mushroom <i>Lentinula edodes</i> - wall glucan
(1,3),(1,6)- β -glucans („branch on branch“ structure)	- yeast <i>Saccharomyces cerevisiae</i> - cell wall glucan - mushroom <i>Schizophyllum commune</i> - wall glucan
(1,3),(1,4)- β -glucans (linear)	- cereal β -glucans - Iceland moss <i>Cetraria islandica</i> - lichenin
(1,3),(1,4)- β -glucans (linear with (1,4)-linked β -glucosyl side branches)	- oyster mushroom (<i>Pleurotus ostreatus</i>) - wall glucan

Anti-inflammatory activity





Enzymatic refining of polysaccharides from Brown macroalgae

Alginate

A polyuronate

Aim: cell free synthesis of a platform chemical, KDG

Alginate degradation

Matis has developed and expressed in high yield thermophilic alginate lyases for near complete degradation of alginate to unsaturated mono-uroantes/DEH

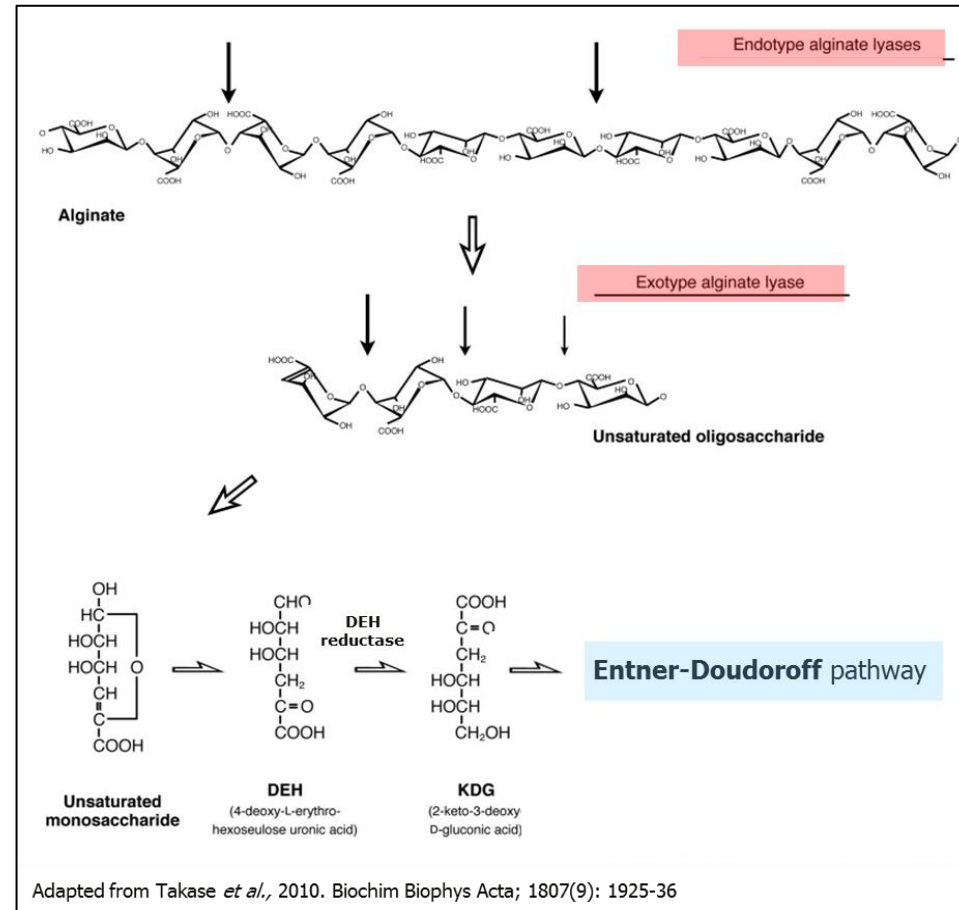
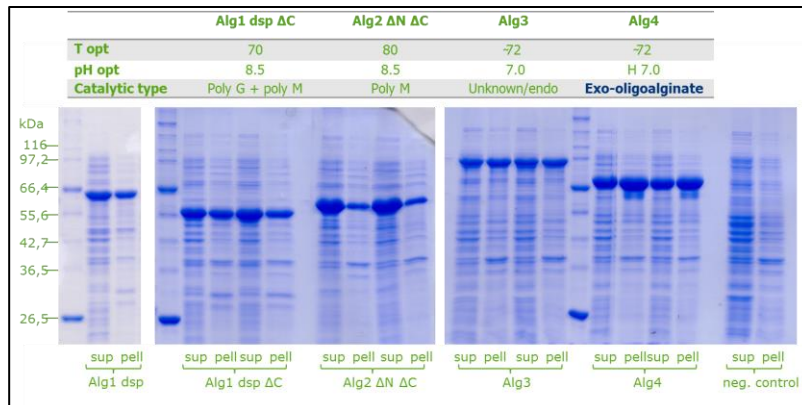
Available in different hosts

E. coli

B. subtilis

Lactobacillus reuterii

For preprocessing of macroalgal biomass

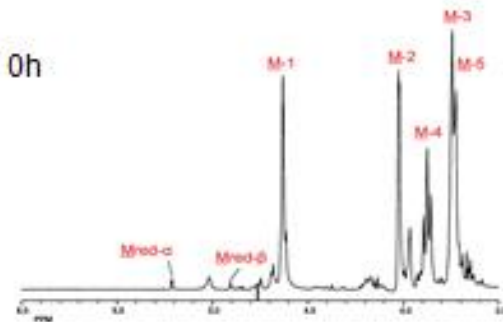


Alg3: an endo-type alginate lyase

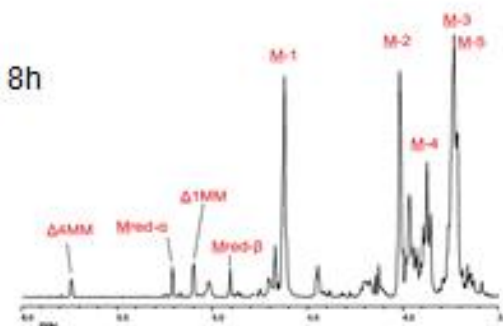
Figure 17. Degradation pattern of AlgRm3 on M-block

1D ¹H NMR

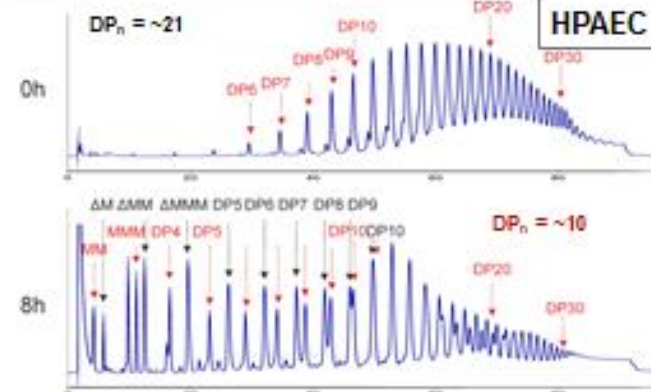
0h



8h



HPAEC



TLC



random activity

yielding these major products:

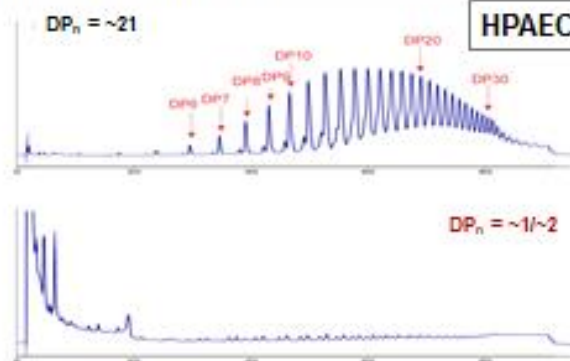
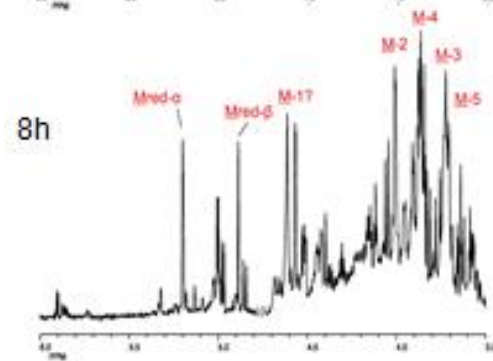
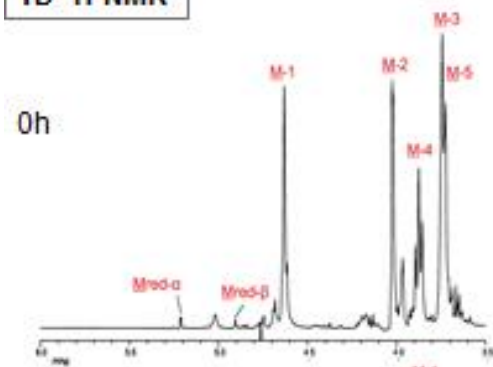
~DP3 ~7 (ΔM..etc.)

...M↓GMMM↓GGG↓GG↓GGGM↓MMMM↓MM↓GM↓GM↓GM↓GM↓MMM↓MMMM...

Alg4: an exo-type alginate lyase – oligo-alginate lyase

Figure 20. Degradation pattern of AlgRm4 on M-block

1D ¹H NMR

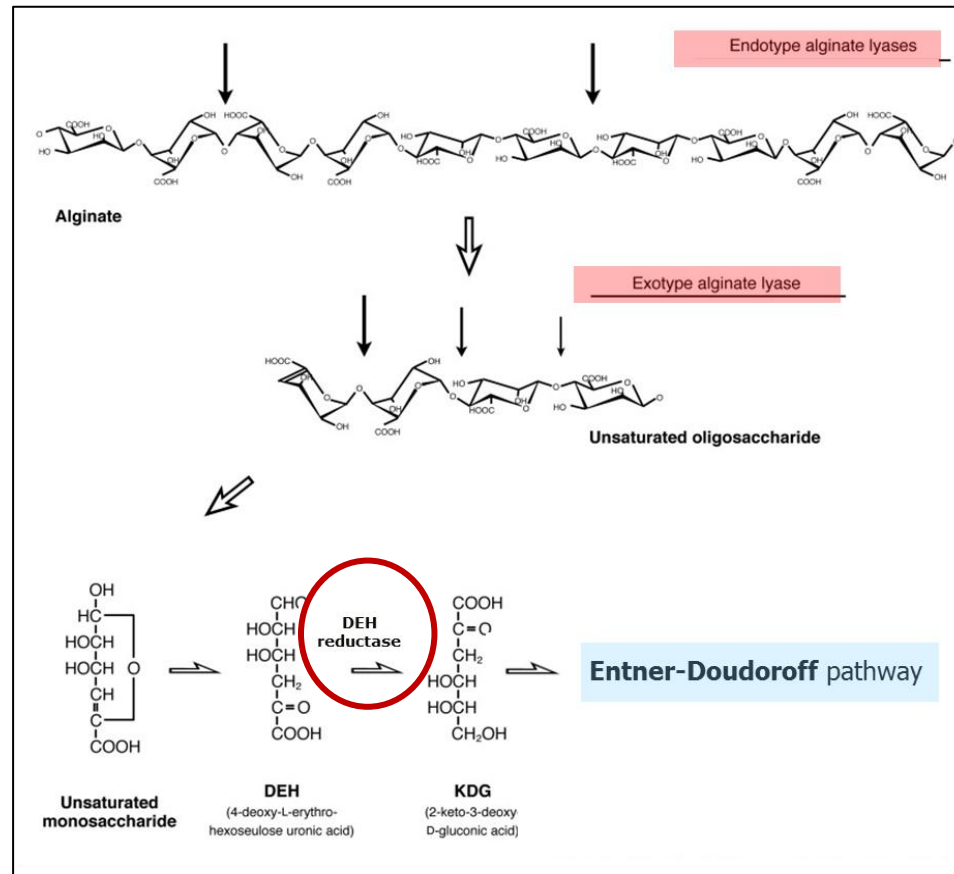


Complete hydrolysis
into monomers:
Δ and Mα/β

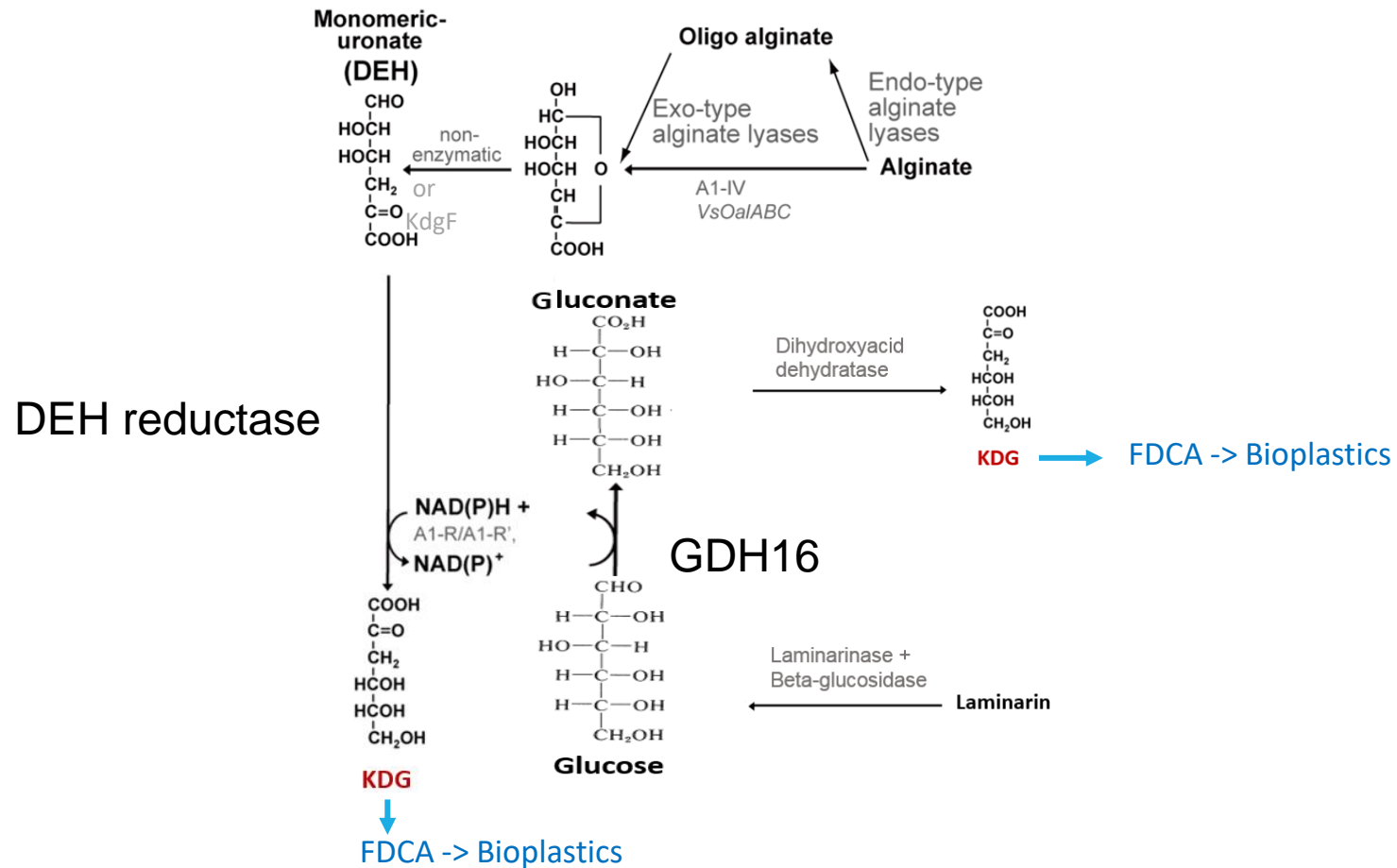
Extra signals observed
at δ 7.34 and δ 8.36 in
1D ¹H NMR spectrum

KDG° from aginate only

- Cell free synthesis of platform chemicals and building blocks from metabolites of alginate catabolism

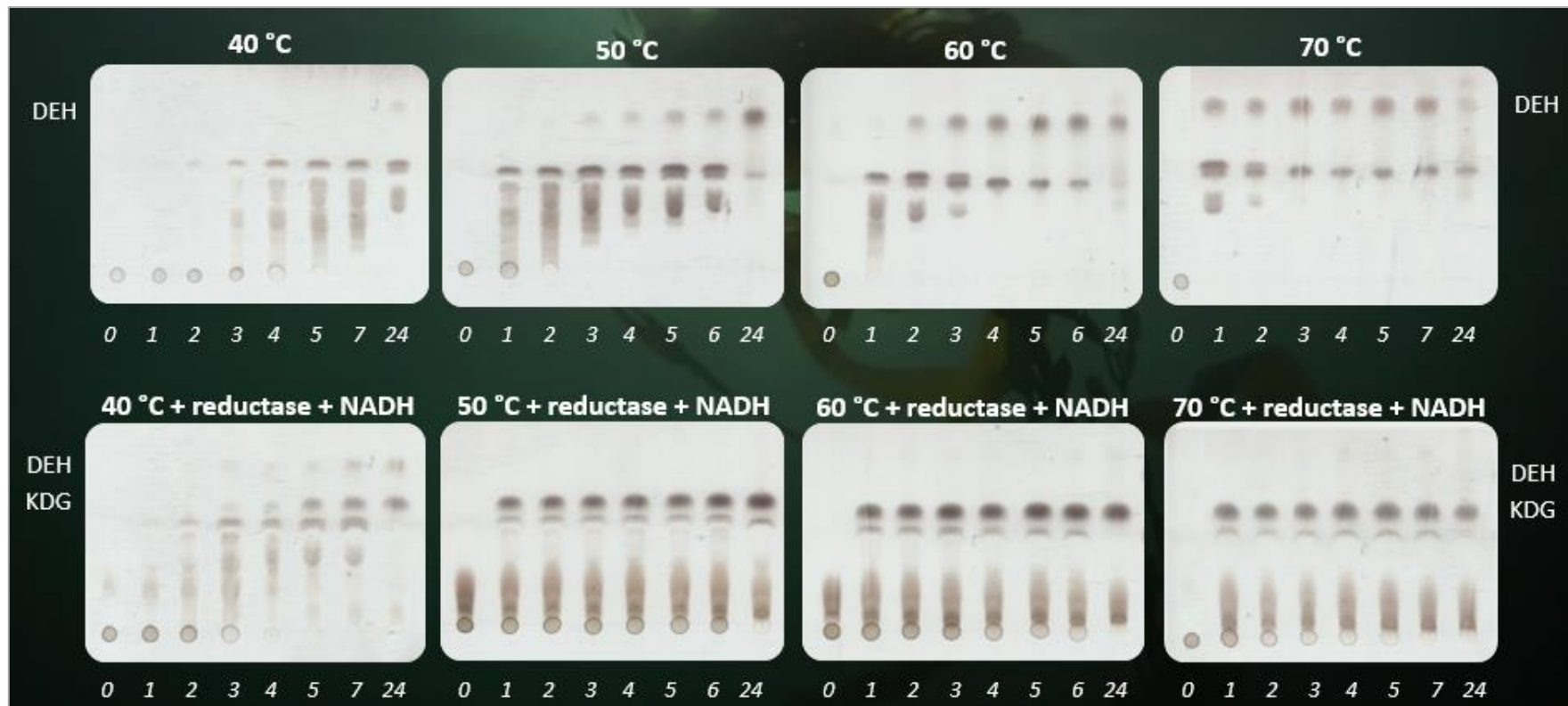


One-pot synthesis of KDG from alginate and laminarin



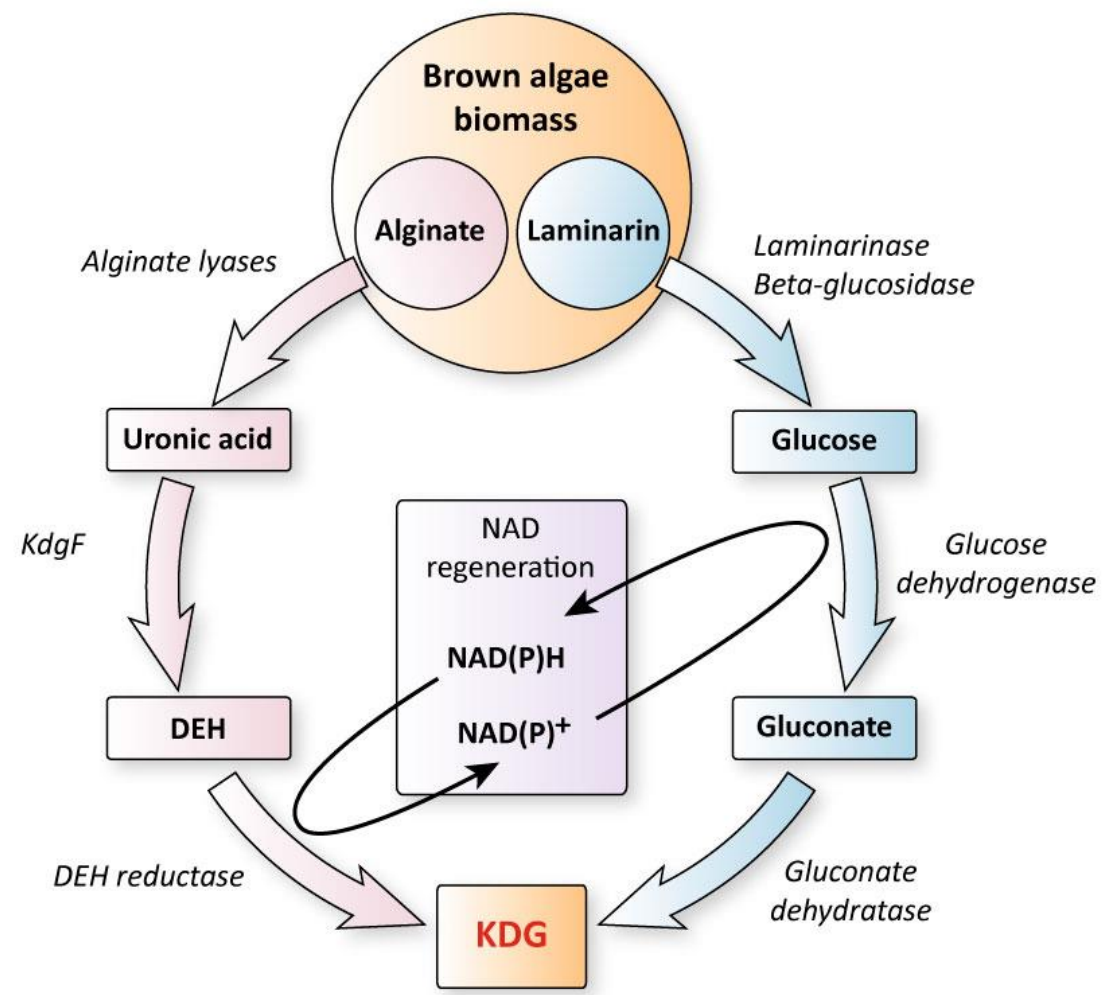
Temperature range

Alginate digestion (top row): **DEH production**, bottom row **KDG production**



Alginate digestion and DEH reduction (bottom row): **KDG production**

Alg3, Alg4 and Adh51 (DEH reductase)





- Thank you for your attention

Coworkers

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Wagenigen Univeristy

Dr. Frits van der Klis

Dr. Paulien Harmsen

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Lund University

Prof. Eva Nordberg Karlsson

References

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<https://www.macrocascade.eu/>